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Synthesis of 1-*N*-[(2*S*,4*S*)- and (2*S*,4*R*)-5-amino-4-fluoro-2-hydroxypentanoyl]dibekacins (study on structure–toxicity relationships)

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Abstract

(2S,4S)- and (2S,4R)-5-azido-2-O-benzyl-4-fluoro-2-hydroxypentanoic acids (15 and 19) have been prepared from L-malic acid (1), and coupled to the H_2N -1 group of 3,2',6'-tris(N-benzyloxy-carbonyl)-3"-N-(trifluoroacetyl)dibekacin (23), to give, after reduction and deblocking, 1-N-[(2S,4S)- and (2S,4R)-5-amino-4-fluoro-2-hydroxypentanoyl]dibekacins (26 and 27). The fluorinated arbekacin analogs showed almost the same antibacterial activities as that of arbekacin, but lower toxicity. Comparision of the toxicity between 26 (and 27) and the arbekacin analogs (28–30) with change of the 1N-side-chain indicates that the observed decrease in toxicity was a function of the chain length rather than the introduction of flourine. © 1998 Elsevier Science Ltd. All rights reserved

Keywords: 5-Amino-4-fluoro-2-hydroxypentanoic acid; Arbekacin; Fluorination; Toxicity

1. Introduction

In a previous paper [1] we reported the synthesis of 1-N-[(2R,3R)- and (2R,3S)-4-amino-3-fluoro-2-hydroxybutanoyl] analogs of arbekacin $\{1-N-[(2S)-4-$ amino-2-hydroxybutanoyl](AHB)dibekacin $\}$ [2] in the hope of obtaining derivatives with decreased toxicity. This was based on the hypothesis that decreasing the basicity of the H_2N-4''' group in arbekacin by introducing flourine, an electron-withdrawing atom, proximal to the amino group (in this case, at C-3''') would decrease the toxicity of arbekacin, as observed in other fluorine-

containing analogs of kanamycins [3–5]. However, contrary to our expectation, the results showed that no such decrease of toxicity occurred. This was explained [1] on the basis of a low decrease in basicity of the H_2N-4''' group compared to that of arbekacin [pKa 10.2 (arbekacin) \rightarrow 8.7]; in other words, the Δ pKa value (=1.5) did not reach 2.2 (pKa 8.0), the value expected to show a toxicity-decreasing effect under physiological conditions (pH \sim 7.0). The present study was designed to examine this concept [1], and describes the synthesis of dibekacin analogs containing 1-N-[(2S,4S)- and (2S,4R)-5-amino-4-fluoro-2-hydroxy-pentanoyl] residues which, compared to AHB, have a one-carbon extended chain.

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Scheme 1.

2. Results and discussion

Synthesis.—First of all, preparation of (2S,4R) and 2S,4S)-5-amino-4-fluoro-2-hydroxypentanoic acids was designed according to the retrosynthesis shown (Scheme 1). After the vicinal carboxyl and hydroxyl groups of L-malic acid (1) had been doubly protected with chloral [6,7], the monoacid 2 was refluxed with thionyl chloride, giving the corresponding acid chloride 3. One-carbon elongation was performed by treatment of 3 with diazomethane to afford an intermediary diazoketone (R-COC⁻HN₂⁺), which was then treated with

ethereal HCl to give the chloromethyl ketone 4. Attempted displacement of the terminal chlorine in 4 by an azide group by NaN₃ in DMF (at room temperature) or in hot CH₃CN gave the corresponding azidoketone 5 in only a trace amount. However, treatment of 4 with NaN₃–Et₄NBr in CH₂Cl₂–CH₃CN, gave 5 in high yield. If, in this reaction, CH₂Cl₂ was omitted, 5 was produced in only a low yield; the reason is unknown. Next, conversion of 5 into the corresponding 2'-ol was attempted with the N₃ group remaining intact. As the azido group is known to be comparatively stable to NaBH₄ [8–10], compound 5 was initially

treated with NaBH₄ (NaBH₃CN or Me₄NBH₄) in MeOH (or THF), but a considerable amount of undesirable by-products (ninhydrin-negative) were produced. Next, Zn(BH₄)₂ in Et₂O was tried, whereupon the oxo group was selectively reduced with simultaneous cyclization, giving a mixture of Υ-lactones (6 (major) and 7). The chirality of 6 and 7 at C-4 was determined by NOE experiments; in 6 irradiation of the higher-field half of the H-5 protons caused a signal increase of H-2 together with the H-3 (H-3a and 3b) protons, indicating the

R-configuration at C-4. In 7, the S configuration at C-4 was deduced from a ROESY experiment for 10, because of the appearance, in 7, of the H-2 and 4 signals at closely proximal positions (Table 1). Benzylation of the 2-hydroxyl group of 6 and 7 was performed by a conventional method (PhCH₂Br in DMF in the presence of Ag₂O), whereupon, however, partial epimerization occurred at C-2, giving a mixture of 2-O-benzyl products (8 and 9 from 6; 10 and 11 from 7) with the 2R isomers predominating. Each of the mixtures was separated

Table 1 ¹H and ¹⁹F NMR data for compounds 3–13, 15, and 17–19 in CDCl₃

Compound	Chemical shifts in ppm $(J \text{ in Hz})$								
	H-2	H-3a	H-3b	H-4	H-5a	H-5b	F-4	$PhCH_2$	Other signals
3	5.91 d J _{2,5} 2	_	_	_	4.88 dt J _{5,1'a} 4	_	_	_	3.52 dd (H-1'a) 3.62 dd (H-1'b)
4	5.90 d J _{2,5} 2	_	_	_	$J_{5,1'b}$ 4 4.89 dt $J_{5,1'a}$ 3.5 $J_{5,1'b}$ 4	_	_	_	$J_{1'a,1'b}$ 19 3.23 dd (H-1'a) 3.47 dd (H-1'b) $J_{1'a,1'b}$ 19
5	5.91 d J _{2,5} 2	_	_	_	4.88 dt $J_{5,1'a}$ 3.5 $J_{5,1'b}$ 4	_	_	_	4.14 s (H-3'a,3'b) 3.10 dd (H-1'a) 3.29 dd (H-1'b) $J_{1'a,1'b}$ 19
6	$J_{2,3a} = 8$ $J_{2,3b} = 9$	2.37 ddd $J_{3a,3b}$ 13.5 $J_{3a,4}$ 8.5	$J_{3b,4}$ 3.5	4.77 dq $J_{4,5a}$ 4 $J_{4,5b}$ 3.5	3.49 dd $J_{5a,5b}$ 13	3.69 dd	_	_	4.05 s (H-3'a,3'b) 2.68 br s (OH)
7	4.63 dd $J_{2,3a} 10.5$ $J_{2,3b} 8.5$	2.09 dt $J_{3a,3b}$ 13 $J_{3a,4}$ 10.5	$2.68 \mathrm{ddd}$ $J_{\mathrm{3b,4}} 6$	4.57 ddt $J_{4,5a}$ 6 $J_{4,5b}$ 3.5	$_{J_{5a,5b}}^{3.51}$ dd $_{J_{5a,5b}}^{3.51}$ 13.5	3.62 dd	_	_	3.87 br s (OH)
8 and 11	$J_{2,3b}$ 6.5 4.27 dd $J_{2,3a}$ 7.5 $J_{2,3b}$ 5.5	$J_{3a,4}$ 10.5 2.28 ddd $J_{3a,3b}$ 13.5 $J_{3a,4}$ 6	$2.33 \text{ddd} \ J_{3b,4} 7$	$J_{4,5b}$ 3.5 4.76 dddd $J_{4,5a}$ 4 $J_{4,5b}$ 3.5	3.42 dd $J_{5a,5b}$ 13.5	3.63 dd	_	4.69 d 4.93 d $J_{\text{gem}} 11.5$	
9 and 10	4.26 dd $J_{2,3a}$ 9	2.10 dt $J_{3a,3b}$ 13	$2.52 \text{ddd} \ J_{3b,4} 6$	$J_{4,5a}$ 6 $J_{4,5b}$ 4.5	3.49 dd $J_{5a,5b}$ 13	3.54 dd	_	4.75 d 4.97 d	
12	$J_{2,3b}$ 8.5 4.17 dd $J_{2,3a}$ 5	$J_{3a,4}$ 9 1.96 ddd $J_{3a,3b}$ 14.5	1.99 dt $J_{3b,4}$ 7.5	$J_{4,5a}$ 6	$_{J_{5a,5b}}^{3.23}$ dd $_{J_{5a,5b}}^{3.25}$ 12.5	3.26 dd	_	J _{gem} 12 4.44 d 4.77 d	2.97 d (OH) $J_{4,\text{OH}}$ 2.5
13	$J_{2,3b}$ 7.5 4.19 dd $J_{2,3a}$ 11 $J_{2,3b}$ 3	$J_{3a,4}$ 4 1.90 dddd $J_{3a,3b}$ 15 $J_{3a,4}$ 2.5	$J_{3b,4}$ 10 $J_{3b,F}$ 13	$J_{4,5b}$ 5 4.90 dddt $J_{4,5a}$ 6 $J_{4,5b}$ 3 $J_{4,F}$ 49	$3.37 \mathrm{ddd} \ J_{5\mathrm{a},5\mathrm{b}} 13.5 \ J_{5\mathrm{a},\mathrm{F}} 24$	3.46 ddd $J_{5b,F}$ 24	-188.4 dddt	J_{gem} 11 4.45 d 4.75 d J_{gem} 11	3.77 s (CO ₂ Me) 3.77 s (CO ₂ Me)
15	4.23 dd $J_{2,3a}$ 11 $J_{2,3b}$ 3	$J_{3a,F}$ 35 1.93 dddd $J_{3a,3b}$ 15 $J_{3a,4}$ 2.5 $J_{3a,F}$ 35	2.28 dddd $J_{3b,4}$ 10 $J_{3b,F}$ 12.5	4.89 dddt J _{4,5a} 6 J _{4,5b} 3 J _{4,F} 49	3.39 ddd $J_{5a,5b}$ 13.5 $J_{5a,F}$ 23.5	$3.46 \mathrm{ddd}$ $J_{5\mathrm{b,F}} 2$	-188.6 dddt	$4.51 d$ $4.79 d$ $J_{\text{gem}} 11$	
17	$J_{2,3a}$ 8.5 $J_{2,3b}$ 4	1.87 ddd $J_{3a,3b}$ 14.5 $J_{3a,4}$ 3	1.94 ddd $J_{3b,4}$ 9.5	3.98 m $J_{4,5a} 6.5$ $J_{4.5b} 4$	3.24 dd $J_{5a,5b}$ 12.5	3.32 dd		4.43 d 4.77 d J_{gem} 11	2.42 d (OH) $J_{4,\text{OH}}$ 4 3.77 s (CO ₂ Me)
18	$J_{2,3a}$ 5.5 $J_{2,3b}$ 6	2.10 ddt $J_{3a,3b}$ 14.5 $J_{3a,4}$ 5.5 $J_{3a,F}$ 25	2.26 dddd $J_{3b,4}$ 7.5 $J_{3b,F}$ 17	$J_{4,5a}$ 6 $J_{4,5a}$ 6 $J_{4,5b}$ 3 $J_{4,F}$ 48	3.35 ddd $J_{5a,5b}$ 13.5 $J_{5a,F}$ 23.5	3.38 ddd $J_{5b,F}$ 25	-185.8 ddq	4.43 d 4.76 d J_{gem} 11.5	$3.78 \text{ s } (\text{CO}_2Me)$
19	$J_{2,3a}$ 6 $J_{2,3b}$ 6	2.13 dddd $J_{3a,3b}$ 15 $J_{3a,4}$ 4.5 $J_{3a,F}$ 27	2.32 ddt $J_{3b,4}$ 8 $J_{3b,F}$ 15	4.93 dddd J _{4,5a} 6 J _{4,5b} 3.5 J _{4,F} 48	3.36 ddd $J_{5a,5b}$ 13.5 $J_{5a,F}$ 23.5	3.39 ddd $J_{5b,F}$ 24.5	-186.0 dddt	4.51 d 4.79 d J_{gem} 11.5	

chromatographically. The structures were confirmed by their ${}^{1}H$ NMR spectra (Table 1) as well as by changes in the specific rotations: a change of $6\rightarrow 8$ or $7\rightarrow 10$ gave a levorotatory change, whereas a change of $6\rightarrow 9$ or $7\rightarrow 11$ gave a large dextrorotatory change. It is noteworthy that 8 and 11, and 9 and 10, showed identical ${}^{1}H$ NMR signals and had opposite rotations. Next, to free the HO-4

group, **8** and **10** were each treated with methanolic HCl, whereupon the desired open-chain esters **12** and **17** were obtained in high yields as syrups (however, long storage or further purification caused partial relactonization, giving the starting materials). Fluorination of **12** or **17** with diethylaminosulfur trifluoride (DAST) [11] gave the corresponding 4-fluoro derivatives (**13** and **18**) with

Table 2 13 C NMR chemical shifts (δ , ppm) and coupling constants ($J_{C,F}$, Hz) for compounds 6–13, 15, 17–19, 21, and 22 in CDCl₃

Compound	C-1	C-2	C-3	C-4	C-5	C-6	Other signals
6	177.3	67.0	33.0	76.0	54.2		
7	176.4	68.1	33.5	75.1	53.6		
8 and 11	173.9	72.9	32.5	76.2	54.0		72.3 ^a
9 and 10	173.8	72.9	32.3	75.0	53.9		72.4 ^a
12	172.4	76.4	36.9	69.1	56.4		52.2 ^b 72.8 ^a
13	172.5	74.0 d (3.7)	35.7 d (21.0)	89.0 d (174.2)	54.4 d (21.0)		52.2 ^b 73.1 ^a
15	176.9	73.7 d (3.7)	35.5 d (21.1)	88.8 d (174.5)	54.3 d (21.0)		73.4 ^a
17	172.8	75.1	36.9	67.6	56.8		52.1 ^b 72.9 ^a
18	172.1	73.7 d (5.6)	35.3 d (21.5)	89.3 d (173.2)	54.1 d (21.3)		52.2 ^b 72.5 ^a
19 21 22	176.1 d (3.0)	73.3 d (4.9) 171.3 171.6	35.0 d (21.2) 71.1 d (5.6) 70.9 d (6.3)	89.1 d (173.6) 33.8 d (20.9) 34.5 d (22.4)	54.0 d (21.5) 85.1 d (173.2) 84.2 d (177.3)	46.4 d (24.3) 45.5 d (26.2)	72.8 ^a 73.5 ^a 72.7 ^a

^{a,b} Signals for PhCH₂ and COOCH₃, respectively.

inversion, accompanied by a small amount of 4eno compound 14 (this was a mixture of E and Z isomers). Alkaline hydrolysis of 13 (or 18) gave the respective free acids 15 (or 19). To ascertain the structures of 15 and 19, they were each reduced catalytically and the amino acids obtained (16 and **20**) were cyclized according to the procedure of Pellegata and coworkers [12], with 1,1,1,3,3,3-hexamethyldisilazane to give the corresponding δ -lactams (21 and 22). The structures were confirmed by ¹H NMR spectra and NOE experiments: in 21, large coupling constants $J_{3,4ax}$, $J_{4ax,F}$, and $J_{6ax,F}$ indicate that BnO-3 (equatorial) and F-5 (axial) are in opposite faces, and this was supported by the NOE experiments: irradiation of F-5 caused increases of the H-3ax and H-4eq signals. A 4H_5 (or 4S_5) structure was proposed as the most probable conformation for 21. In 22, a structure with the BnO-3 and F-5 in the same face was confirmed on account of the observation of an NOE between H-3 and H-5, although the correct conformation is not clear. These results indicate that 15 and 19 have the expected structures (Table 2).

Coupling of 15 (or 19) with 3.2',6'-tris[N-benzyloxycarbonyl(=Z)-3''-N-trifluoroacetyl]dibekacin 23 having the H_2N -1 group free was performed utilizing the active ester 24 (or 25) of 15 (or 19) according to conventional methodology [1,13]. The condensed 1-N-acyl derivatives were successively de(trifluoroacetyl)ated, reduced (N_3 -5'''), and hydrogenolyzed (three Z groups) to give the final products (26 and 27).

Biological activity.—Antibacterial activities of 26 and 27 as compared with arbekacin (Table 4)

indicate that both compounds had activities almost identical to that of arbekacin. This indicates that the activity is not influenced by the one-carbon elongation or the orientation of F-4". It is noteworthy that the more-active counterpart, 1-N-[(2R,3R)-[not

Table 3 13 C NMR chemical shifts (δ , ppm) and coupling constants ($J_{C,F}$, Hz) for compounds **26–30** and arbekacin (ABK) in DCl–D₂O (pD 3)

220 (P2	2)					
	26	27	28	29	30	ABKa
C-1	49.6	49.7	49.5	49.6	49.6	49.6
C-2	31.2	31.1	31.1	31.2	31.3	31.2
C-3	49.8	49.8	49.9	49.9	49.9	49.8
C-4	78.6	78.6	78.6	78.6	78.6	78.5
C-5	75.7	75.7	75.8	75.7	75.7	75.7
C-6	81.0	80.7	81.7	80.9	80.9	81.1
C-1'	96.0	96.0	96.0	96.0	95.9	95.9
C-2'	49.7	49.7	49.7	49.7	49.8	49.7
C-3'	21.4	21.4	21.5	21.5	21.5	21.5
C-4'	26.2	26.2	26.3	26.3	26.3	26.3
C-5'	66.9	66.8	66.9	66.9	66.9	66.9
C-6'	43.5	43.4	43.5	43.5	43.6	43.5
C-1"	98.9	98.7	99.2	98.8	98.7	98.9
C-2"	68.8	68.8	68.9	68.9	68.9	68.8
C-3"	56.0	56.0	56.1	56.1	56.1	56.1
C-4"	66.4	66.4	66.5	66.5	66.5	66.5
C-5"	73.0	73.0	73.1	73.1	73.1	73.0
C-6"	60.7	60.7	60.8	60.8	60.8	60.7
C-1′′′	176.6	176.4	173.8	177.1	177.5	176.3
C-2′′′	68.2d	68.9 d	68.7	71.8	72.1	70.5
C-3′′′	37.0 d	36.8 d	42.9	31.3	33.9	31.7
C-4′′′	88.4 d	89.4 d		24.1	22.8	37.9
C-5′′′	43.9 d	43.7 d		40.1	27.3	
C-6′′′					40.3	
$J_{C-2''',F}$	3.5	\sim 4				
$J_{ ext{C-3}''', ext{F}}$	19.5	19.4				
$J_{ ext{C-4}''', ext{F}}$	170.4	169.6				
$J_{ ext{C-5}''', ext{F}}$	20.3	19.6				

^a Ref. [1].

(2R,3S)]-4-amino-3-fluoro-2-hydroxybutanoyl]dibekacin reported previously [1] has an F-3-D structure. In terms of acute toxicity, both **26** (LD₅₀~130 mg/kg, mice, intravenous injection) and **27** (LD₅₀~125) showed almost the same value, but this was considerably lower than that of arbekacin (LD₅₀~75). This indicates that antibacterial activity and toxicity do not necessarily change in parallel [4]. This lower toxicity of **26** and **27** as compared with arbekacin, however, is not to be ascribed to the decrease in basicity of H₂N-5" that results from the induction of the F-4" atom, as had

been predicted in our previous work and was explained in detail in [1] (for example, arbekacin and an arbekacin analog having the (2R,3R)-4-amino-3-fluoro-2-hydroxybutanoyl residue have similar toxicities). This unexpected decrease in toxicity in the present derivatives **26** and **27** must, therefore, be ascribed to the difference in the chainlength or the position of the fluorine atom introduced.

To clarify the problem, three arbekacin analogs with varying lengths of the side chain, without fluorine, were conventionally prepared, namely,

Table 4 Minimal inhibitory concentration^a (μ g/mL) of compounds 26–30 and arbekacin (ABK)

Test organism ^b	26	27	28	29	30	ABK
St. a. FDA 209 P	0.10	0.10	0.39	0.10	0.20	0.20
St. a. Smith	< 0.05	< 0.05	0.05	< 0.05	0.10	0.05
St. a. Ap01 ^c	> 100	> 100	> 100	> 100	> 100	> 100
Micr. l. FDA16	3.13	1.56	3.13	6.25	> 100	3.13
Micr. l. PCI 1001	3.13	1.56	3.13	6.25	> 100	3.13
B. c. ATCC 10702	0.78	0.78	1.56	1.56	1.56	1.56
Coryn. b. 1810	1.56	0.39	0.78	3.13	50	0.39
E. c. NIHJ	0.39	0.39	0.39	0.39	1.56	0.39
E. c. K-12 R5 ^d	> 100	> 100	> 100	> 100	> 100	> 100
E. c. K-12 ML1629 ^e	0.78	1.56	1.56	1.56	6.25	1.56
E. c. K-12 ML1410 R81e	1.56	1.56	1.56	1.56	25	1.56
E. c. K-12 LA290 R55 ^f	3.13	3.13	1.56	0.78	3.13	1.56
E. c. K-12 LA290 R64	0.39	0.39	0.39	0.39	1.56	0.39
E. c. W677	0.39	0.39	0.39	0.39	1.56	0.39
E. c. JR66/W677 ^{f,g}	1.56	1.56	1.56	1.56	6.25	1.56
E. c. JR225 ^h	0.78	1.56	0.78	0.78	3.13	0.78
Kl. p. PCI602	0.39	0.78	1.56	0.78	3.13	0.78
Kl. p. 22#3038 ^{f,g}	0.78	0.78	0.78	0.78	3.13	0.78
Sh. s. JS11746	0.78	0.78	0.78	0.78	3.13	0.78
Sal. e. 1891	3.13	3.13	3.13	6.25	50	3.13
Serr. marc.	25	3.13	3.13	12.5	100	6.25
Prot. r. GN311	1.56	3.13	3.13	1.56	3.13	6.25
<i>Prov.</i> sp. Pv 16 ⁱ	6.25	1.56	3.13	6.25	50	1.56
<i>Prov.</i> sp. 2991 ⁱ	25	12.5	25	25	> 100	6.25
Ps. aerug. A3	0.39	0.39	0.78	0.39	0.78	0.39
Ps. aerug. H9 ^g	1.56	1.56	1.56	1.56	6.25	1.56
Ps. aerug. GN315 ^d	25	6.25	6.25	25	> 100	6.25

^a Judged by the agar dilution streak method (Mueller–Hinton agar, 37 °C, 18 h).

1-N-[(2S)-3-amino-2-hydroxypropanoyl]dibekacin (28) [14], 1-N-[(2S)-5-amino-2-hydroxypentanoyl]dibekacin (29), and 1-N-[(2S)-6-amino-2-hydroxyhexanoyl]dibekacin (30) (Table 3). The antibacterial activities (Table 4) of compounds 28, 29, and arbekacin were found to be almost the same, whereas 30 was less active. However, it was established that the acute toxicity of these compounds was significantly influenced by the chain-length, with decrease in toxicity with increase in length (28: \sim 65, **29**: \sim 120, **30**: \sim 125 mg/kg). A specific feature is that 26, 27, and 29 with the same chain-length, had analogous toxicities, indicating that the decrease in toxicity of 26 (and 27) compared to arbekacin must be ascribed to the difference in chain length, and not to the introduction of fluorine at C-4'''.

3. Experimental

General methods.—Melting points were determined on a Kofler block and are uncorrected. Optical rotations were determined with a Perkin–Elmer 241 polarimeter. IR spectra were measured with a Jasco A-202 grating spectrophotometer. ¹H, ¹³C, and ¹⁹F NMR spectra were recorded at 250 (¹H, for 3–5 and 7) and 235.3 MHz (¹⁹F) with a Bruker WM 250 spectrometer, and at 500 (¹H), 125.8 (¹³C), and 470.6 MHz (¹⁹F, for 21) with an AMX 500 spectrometer. Chemical shifts (δ) of ¹H, ¹³C, and ¹⁹F spectra were measured downfield from internal Me₄Si (for ¹H and ¹³C) and internal Freon 11 (for ¹⁹F), unless otherwise stated, and were confirmed, when necessary, by shift-correlated 2D spectra. Thin-layer chromatography (TLC) was

b Abbreviations: St. a., Staphylococcus aureus; Micr. l., Micrococcus luteus; B. c., Bacillus cereus; Coryn. b., Corynebacterium bovis; E. c., Escherichia coli; Kl. p., Klebsiella pneumoniae; Sh. s., Shigella sonnei; Sal. e., Salmonella enteritidis; Serr. mar., Serratia marcescens; Prot. r., Proteus rettgeri; Prov., Providencia; Ps. aerug., Pseudomonas aeruginosa.

^c Resistant strain producing AAD(4').

d AAC(6').

e APH(3')-I.

f AAD(2").

^g APH(3')-II.

^h AAC(3).

i AAC(2').

performed on Kieselgel 60 F₂₅₄ (Merck), and column chromatography, on Wakogel C-200, unless stated otherwise. HPLC was performed on a C.I.G. pre-packed silica-gel column (Kusano Kagakukikai Co., Japan) using 1:3 hexane–EtOAc.

(2R,5S) - 5 - (2 - Chloro - 2 - oxoethyl) - 2 - trichloro-methyl-1,3-dioxolan-4-one (3).—A mixture of **2** [6,7] [11.9 g, 45 mmol; $[\alpha]_D^{23} + 38.5^\circ$ (c 5, EtOH), $[\alpha]_D^{23} + 35^\circ$ (c 1, CHCl₃); lit [6], $[\alpha]_D^{28} + 39.1^\circ$ (c 5.14, EtOH), lit [7], $[\alpha]_D^{20} + 33.03$ (c 1.005, CHCl₃)] and SOCl₂ (20 mL, 270 mmol) was gently refluxed for 60 h. Evaporation of SOCl₂ gave a residue that crystallized from hexane to give **3** as needles (11.8 g, 93%); mp 74–75 °C (lit [6] 70–72 °C), $[\alpha]_D^{24} + 24^\circ$ (c 1, CHCl₃); IR (KBr): 1780 (C=O), 1820 cm⁻¹ [C=O (lactone)]. Anal. Calcd for C₆H₄Cl₄O₄: C, 25.56; H, 1.43; Cl, 50.30. Found: C, 25.79; H, 1.50; Cl, 50.19.

(2R,5S)-5-(3-Chloro-2-oxopropyl)-2-trichloromethyl-1,3-dioxolan-4-one (4).—To a solution of 3 $(10.0\,\mathrm{g},\ 35\,\mathrm{mmol})$ in $\mathrm{Et_2O}$ $(100\,\mathrm{mL})$ was added $0.25 \,\mathrm{M}$ ethereal CH₂N₂ (280 mL, 70 mmol), and the solution was kept for 30 min at room temperature. Ethereal 1.2 M HCl (70 mL) was added, and the mixture was kept for a further 30 min. TLC (6:1 toluene–EtOAc) of the solution showed a spot at R_f 0.55. After concentration, the residue was chromatographed (6:1 toluene-EtOAc) to give 4 as a crystalline solid (9.8 g, 93%); mp 75–76°C (toluene–hexane), $\left[\alpha\right]_{D}^{23} + 33^{\circ}$ (c 1, CHCl₃); IR (KBr): 1735 (C=O), 1815 cm⁻¹ [C=O (lactone)]; ¹³C NMR (CDCl₃; confirmed by the ¹H-¹³C HMBC): δ 198.8 (C-2'), 170.3 (C-4), 105.2 (C-2), 97.7 (CCl₃), 70.5 (C-5), 47.5 (C-3'), 40.7 (C-1'). Anal. Calcd for C₇H₆Cl₄O₄: C, 28.41; H, 2.04; Cl, 47.92. Found: C, 28.67; H, 1.87; Cl, 47.98.

(2R,5S)-5-(3-Azido-2-oxopropyl)-2-trichloromethyl-1,3-dioxolan-4-one (5).—To a cold $(0^{\circ}C)$ solution of 4 (6.0 g, 20 mmol) in 6:5 CH₂Cl₂-CH₃CN (110 mL) were added NaN₃ (1.7 g, 26 mmol) and Et₄NBr (4.5 g, 21 mmol), and the mixture was stirred for 3h in the cold; TLC (6:1 toluene–EtOAc) of the organic layer showed a spot at R_f 0.45. After dilution with CHCl₃ (600 mL), the solution was washed with water, dried (Na₂SO₄), and concentrated. The residue was purified by chromatography (6:1 toluene-EtOAc) to give 5 as a crystalline solid (5.28 g, 86%), mp 105-106 °C (toluene-hexane), $[\alpha]_D^{23} + 38^\circ$ (c 1, CHCl₃); IR (KBr): 1725, 1830, 2110 cm^{-1} (N₃). Anal. Calcd for C₇H₆Cl₃N₃O₄: C, 27.79; H, 2.00; Cl, 35.16; N, 13.89. Found: C, 28.09; H, 2.24; Cl, 35.37; N, 13.71.

*5-Azido-3,5-dideoxy-*L-erythro-*pentono-1,4-lactone* (6) and 5-azido-3,5-dideoxy-D-threo-pentono-1,4lactone (7).—To a cold (-10°C) solution of 5 (1.95 g, 6.45 mmol) in THF (60 mL) was added ~ 0.07 M ethereal Zn(BH₄)₂ (first 30 mL, and then another 30 mL after 1 h; \sim 4 mmol in total), and the mixture was kept for 2h in the cold. TLC (1:1 toluene–EtOAc) of the organic layer showed spots at R_f 0.25 (6), 0.2 (7), and 0 (cf. 5: R_f 0.7). After addition of AcOH (~0.8 mL), the mixture was extracted with EtOAc. The extracts were washed with ag Na₂SO₄ (satd), dried (Na₂SO₄), and concentrated to a syrup. HPLC of the syrup afforded 6 (565 mg, 56%) and 7 (357 mg, 35%) as syrups. Compound 6 had $\left[\alpha\right]_{D}^{22}-137^{\circ}$ (c 1, CHCl₃); IR (neat): 1780 (C=O), 2110 cm^{-1} (N₃); NOE difference spectroscopy: irradiation of H-5a, H-2 (0.4%), H-3a (0.6%), H-3b (1.2%), and H-4 (1.8%) signals were increased. Anal. Calcd for C₅H₇N₃O₃: C, 38.22; H, 4.49; N, 26.74. Found: C, 38.32; H, 4.31; N, 26.41. Compound 7 had $[\alpha]_D^{22}$ $+59^{\circ}$ (c 1, CHCl₃); IR (neat): 1780, 2110 cm⁻¹. Anal. Calcd for C₅H₇N₃O₃: C, 38.22; H, 4.49; N, 26.74. Found: C, 38.23; H, 4.75; N, 26.63.

*5-Azido-2-O-benzyl-3,5-dideoxy-*L-erythro-*pent*ono-1,4-lactone (8) and 5-azido-2-O-benzyl-3,5dideoxy-L-threo-pentono-1,4-lactone (9).—To a solution of 6 (395 mg) in DMF (8 mL) were added Ag₂O (1.16 g) and excess PhCH₂Br (0.6 mL), and the mixture was stirred for 3h at room temperature; TLC (4:1 toluene-EtOAc) of the organic layer showed two spots at R_f 0.55 (8, major) and 0.45 (9) (cf. 6: R_f 0.05). Filtration followed by concentration in vacuo gave a syrup, which was extracted with CHCl₃. The extracts were washed with 0.05 M ag HCl and water, dried (Na₂SO₄), and concentrated. The residue was chromatographed (8:1 toluene-EtOAc) to give 8 (503 mg, 81%) and 9 (68 mg, 11%) as syrups. Compound 8 had $[\alpha]_D^{21}$ -160° (c 1, CHCl₃). Anal. Calcd for C₁₂H₁₃N₃O₃: C, 58.29; H, 5.30; N, 16.99. Found: C, 58.17; H, 5.31; N, 16.75. Compound **9** had $[\alpha]_D^{20}$ -5° (c 1, CHCl₃). Anal. Calcd for C₁₂H₁₃N₃O₃: C, 58.29; H, 5.30; N, 16.99. Found: C, 58.05; H, 5.26; N, 16.77.

5-Azido-2-O-benzyl-3,5-dideoxy-D-threo-pentono-1,4-lactone (10) and 5-azido-2-O-benzyl-3,5-dideoxy-D-erythro-pentono-1,4-lactone (11).—To a solution of 7 (1.08 g) in DMF (22 mL) were added Ag₂O (3.20 g) and excess PhCH₂Br (1.64 mL), and the mixture was stirred for 3 h at room temperature. TLC (4:1 toluene–EtOAc) of the organic layer showed two spots at R_f 0.45 (10, major) and 0.55

(11). The mixture was treated as first described above to give 10 (1.36 g, 80%) and 11 (160 mg, 9%) as syrups. Compound 10 had $[\alpha]_D^{24} + 4^\circ$ (c 1, CHCl₃); the ¹H NMR spectrum was the same as that of 9; in ROESY, a cross peak was observed between H-2 and H-4. Anal. Calcd for $C_{12}H_{13}N_3O_3$: C, 58.29; H, 5.30; N, 16.99. Found: C, 58.54; H, 5.37; N, 17.23. Compound 11 had $[\alpha]_D^{20} + 161^\circ$ (c 1, CHCl₃); the ¹H NMR spectrum was the same as that of 8. Anal. Calcd for $C_{12}H_{13}N_3O_3$: C, 58.29; H, 5.30; N, 16.99. Found: C, 58.00; H, 5.52; N, 16.69.

Methyl (2S,4R)-5-azido-2-O-benzyl-2,4-dihydroxy-pentanoate (12).—A solution of **8** (4.58 g) in 0.1 M methanolic HCl (75 mL) was kept for 10 min at room temperature. TLC (4:1 toluene–EtOAc) of the solution showed two spots at R_f 0.35 (12) and 0.55 (**8**, minor). After addition of CHCl₃ (500 mL), the solution was washed with water, dried (Na₂SO₄), and concentrated. Chromatography (6:1 toluene–EtOAc) of the syrup gave 12 as a syrup (4.25 g, 82%) together with **8** recovered (640 mg). Compound 12 had IR (neat): 1740 (C=O), 2110 cm⁻¹ (N₃).

Methyl (2S,4S)-5-azido-2-O-benzyl-4-fluoro-2hydroxypentanoate (13) and methyl (S)-5-azido-2-O-benzyl-2-hydroxy-4-pentenoate (14).—To a solution of DAST (2.0 mL, 15.1 mmol) in 8:1 benzenepyridine (45 mL) was added a solution of 12 $(1.50\,\mathrm{g},\ 5.37\,\mathrm{mmol})$ in benzene $(30\,\mathrm{mL})$, and the mixture was kept for 1 h at room temperature and then for 2h at 60°C. TLC (10:2:1 cyclohexane-CHCl₃-acetone) of the solution showed spots at R_f 0.45 (14), 0.35 (13, major), 0.2 (trace), and 0.1. The mixture was poured into aq NaHCO3 (satd, 300 mL) and toluene (300 mL), and after shaking for 10 min, the organic layer that separated was washed with water, dried (Na₂SO₄), and concentrated. Chromatography (10:2:1 cyclohexane-CHCl₃-acetone) of the residue gave 13 as a syrup (1.16 g, 77%), along with syrupy **14** (84 mg), ¹H NMR (CDCl₃): (the ratio of the E and Z isomers was $\sim 1:2$) δ 6.23 [dt, 1 H, H-5 (Z)], 5.94 [dt, 1 H, H-5 (E)], 5.34 [dt, 1 H, H-4 (E)], 4.94 [q, 1 H, H-4 (Z)], 4.44 and 4.71 [each d of 1 H, J 12 Hz, PhC H_2 (Z)], 4.43 and 4.72 [each d of 1 H, J 12 Hz, PhC H_2 (E)], 3.98 [dd, 1 H, H-2 (Z)], 3.95 [dd, 1 H, H-2 (E)], 3.75 (s, COOMe), 2.58 [dddd, 1 H, H-3b (Z)], 2.54 [ddt, 1 H, H-3a (Z)], 2.49 [m, 2 H, H-3a,3b (E)]; J (E isomer): $J_{2,3a}$ 7, $J_{2,3b}$ 5.5, $J_{3a,4} \approx J_{3b,4}$ 7.5, $J_{4.5}$ 13.5, $J_{3a.5} \approx J_{3b.5} \sim 1.5$ Hz; (Z isomer): $J_{2.3a}$ 7, $J_{2,3b}$ 5.5, $J_{3a,3b}$ 15, $J_{3a,4} \approx J_{3b,4} \approx J_{4,5}$ 7.5, $J_{3a,5} \approx J_{3b,5}$

1.5 Hz. Compound **13**, $[\alpha]_D^{24}$ -74° (*c* 1, CHCl₃). Anal. Calcd for C₁₃H₁₆FN₃O₃: C, 55.51; H, 5.73; F, 6.75; N, 14.94. Found: C, 55.42; H, 5.81; F, 6.92; N, 14.93.

(2S,4S)-5-Azido-2-O-benzyl-4-fluoro-2-hydroxy-pentanoic acid (15).—To a solution of 13 (1.80 g) in MeOH (36 mL) was added M aq NaOH (9 mL), and the solution was kept for 1 h at room temperature. TLC (3:1 CHCl₃–MeOH) of the solution showed a single spot at R_f 0.5 (cf. 13: R_f 0.9). The solution was concentrated to a low volume, diluted with water, and acidified with M aq HCl to pH ~1 under cooling. Extraction of the product with EtOAc followed by concentration gave 15 as a crystalline solid (1.70 g, 99%), mp 99–100 °C (CHCl₃–hexane), [α]_D²⁴ –81° (c 1, CHCl₃). Anal. Calcd for C₁₂H₁₄FN₃O₃: C, 53.93; H, 5.28; F, 7.11; N, 15.72. Found: C, 53.74; H, 5.18; F, 6.86; N, 15.92.

Methyl (2S,4S)-5-azido-2-O-benzyl-2,4-dihydroxy-pentanoate (17).—A solution of 10 (1.56 g) in 0.1 M methanolic HCl (25.5 mL) was kept for 10 min at room temperature. TLC (4:1 toluene—EtOAc) of the solution showed two spots at R_f 0.4 (17) and 0.45 (10, minor). Processing as described for 12 gave a syrup, chromatography (6:1 toluene—EtOAc) of which gave 17 as a syrup (1.41 g, 80%) together with 10 recovered (230 mg). Compound 17 had IR (neat): 1740 (C=O), 2110 cm⁻¹ (N₃).

Methyl (2S,4R)-5-azido-2-O-benzyl-4-fluoro-2-hydroxypentanoate (18).—To a solution of DAST (1.4 mL) in 8:1 benzene–pyridine (30 mL) was added a solution of 17 (1.00 g) in benzene (20 mL), and the mixture was treated as described for 13. TLC (10:2:1 cyclohexane–CHCl₃–acetone) of the solution showed spots at R_f 0.45 (14), 0.35 (18, major), 0.2 (trace), and 0.1 (a mixture of 17 and several by-products). Similar purification as described for 13 gave 18 as a syrup (545 mg, 54%), together with 14 (125 mg). Compound 18 had $[\alpha]_D^{23}$ –60° (*c* 1, CHCl₃). Anal. Calcd for $C_{13}H_{16}FN_3O_3$: C, 55.51; H, 5.73; F, 6.75; N, 14.94. Found: C, 55.63; H, 5.79; F, 6.98; N, 15.15.

(2S,4R)-5-Azido-2-O-benzyl-4-fluoro-2-hydroxy-pentanoic acid (19).—To a solution of 18 (350 mg) in MeOH (7 mL) was added M aq NaOH (1.8 mL), and the solution was kept for 1 h at room temperature. TLC (3:1 CHCl₃–MeOH) of the solution showed a single spot at R_f 0.5 (cf. 18: R_f 0.9). Similar treatment as described for 15 gave 19 as a syrup (315 mg, 95%). An analytical sample (syrup) was prepared by column chromatography using

the lower phase of 20:1:1 CHCl₃–MeOH–20% aq AcOH, $[\alpha]_D^{22}$ –49° (*c* 1, CHCl₃). Anal. Calcd for C₁₂H₁₄FN₃O₃: C, 53.93; H, 5.28; F, 7.11; N, 15.72. Found: C, 54.15; H, 5.37; F, 7.12; N, 15.87.

(3S,5S) - 3 - Benzyloxy - 5 - fluoro - 2 - piperidinone (21).—A solution of 15 (200 mg) in 3:1 MeOH– H₂O (8 mL) was hydrogenated under H₂ in the presence of Pd-black for 40 min at room temperature. TLC (1:1 CHCl₃-MeOH) of the solution showed a single spot at R_f 0.2 (cf. 15: R_f 0.65). Filtration followed by concentration gave 16 as a pale-yellow solid (168 mg, 93%). A mixture of the solid in CH₃CN (1.7 mL), 1,1,1,3,3,3-hexamethyldisilazane (3.1 mL), and 1.2 M ethereal HCl (0.6 mL; prepared by introducing HCl vapor into Et₂O) was refluxed overnight. TLC (1:3 toluene-EtOAc) of the solution showed a spot at R_f 0.25. Addition of MeOH (8 mL) followed by concentration gave a residue, which was purified by chromatography (1:3 toluene-EtOAc) to give 21 as a crystalline solid (90 mg, 58%), mp 119– 120 °C (toluene-hexane), $\left[\alpha\right]_{D}^{22}$ -115° (c 1, CHCl₃); IR (KBr): 1630 cm⁻¹ (amide); ¹H NMR (CDCl₃): δ 5.95 (br s, 1 H, NH), 5.07 (dddq, 1 H, $J_{5,4ax}$ 3.5, $J_{5,4eq}$ 5, $J_{5,F}$ 48, $J_{5,6ax}$ 4, $J_{5,6eq}$ 3, $J_{5,NH}$ \sim 1 Hz, H-5), 4.72 and 5.02 (each d of 1 H, J 12 Hz, PhC H_2), 4.13 (dd, 1 H, $J_{3,4ax}$ 9, $J_{3,4eq}$ 5.5 Hz, H-3), 3.62 (dddd, 1 H, J_{6ax,5} 4, J_{6ax,F} 34, J_{6ax,6eq} 13.5, $J_{6ax,NH}$ 2 Hz, H-6ax), 3.52 (dddt, 1 H, $J_{6eq,4eq}$ 2, $J_{6eq,5}$ 3, $J_{6eq,F}$ 18, $J_{6eq,6ax}$ 13.5, $J_{6eq,NH}$ 3 Hz, H-6eq), 2.47 (ddddd, 1 H, J_{4eq,3} 5.5, J_{4eq,4ax} 14.5, $J_{4eq,5}$ 5, $J_{4eq,6eq}$ 2, $J_{4eq,F}$ 12.5 Hz, H-4eq), 2.19 (dddd, 1 H, $J_{4ax,3}$ 9, $J_{4ax,4eq}$ 14.5, $J_{4ax,5}$ 3.5, $J_{4ax,F}$ 34 Hz, H-4ax); NOE difference spectroscopy: irradiation of F increased the signals of H-3 (27%), H-4eq (21%), H-5 (100%; taken as the reference for the increases), and H-6eq (23%). ¹⁹F NMR (CDCl₃): δ -182.8 (dddt). Anal. Calcd for C₁₂H₁₄FNO₂: C, 64.56; H, 6.32; F, 8.51; N, 6.28. Found: C, 64.77; H, 6.24; F, 8.30; N, 6.43.

(3S,5R) - 3 - Benzyloxy - 5 - fluoro - 2 - piperidinone (22).—A solution of 19 (150 mg) in 3:1 MeOH–H₂O (6 mL) was hydrogenated in a similar manner as described for 21. TLC (1:1 CHCl₃–MeOH) of the solution showed a single spot at R_f 0.2 (*cf.* 19: R_f 0.65). Post-treatment as described for 16 gave 20 as a pale-yellow solid (128 mg, 95%). The solid was then treated as described for 21 to give 22 as a crystalline solid (73 mg, 62%); mp 85.5–86.5 °C (toluene–hexane), [α]_D²³ –83° (*c* 1, CHCl₃); IR (KBr): 1645 cm⁻¹ (amide); ¹H NMR (CDCl₃): δ 6.09 (br s, 1 H, NH), 4.94 (m, 1 H, $J_{5,4a}$ 5.5, $J_{5,4b}$ 6,

 $J_{5,F}$ 49, $J_{5,6a}$ 4, $J_{5,6b}$ 5 Hz, $J_{5,NH} \le 0.5$, H-5; irradiation of NH caused collapse to ddddd), 4.73 and 4.99 (each d of 1 H, J 12 Hz, PhC H_2), 3.88 (dd, 1 H, $J_{3,4a}$ 9, $J_{3,4b}$ 7 Hz, H-3), 3.54 (ddddd, 1 H, $J_{6b,5}$ 5, $J_{6b,F}$ 14, $J_{6b,6a}$ 13.5, $J_{6b,NH}$ 4 Hz, H-6b), 3.43 (ddddd, 1 H, $J_{6a,4b} \le 1$, $J_{6a,5}$ 4, $J_{6a,6b}$ 13.5, $J_{6a,F}$ 24, $J_{6a,NH}$ 3 Hz, H-6a), 2.52 (ddddd, 1 H, $J_{4b,3}$ 7, $J_{4b,4a}$ 14.5, $J_{4b,5}$ 6, $J_{4b,6a} \le 1$, $J_{4b,F}$ 22 Hz, H-4b), 2.22 (ddddd, 1 H, $J_{4a,3}$ 9, $J_{4a,4b}$ 14.5, $J_{4a,5}$ 5.5, $J_{4a,6b}$ 1, $J_{4a,F}$ 21 Hz, H-4a); NOE difference spectroscopy: irradiation of H-3 increased the signals of H-4b (3.3%), H-5 (1.9%), and H-6a (2.2 %). ¹⁹F NMR (CDCl₃): δ -178.3 (dddq; $J_{F,NH} \sim 1.5$ Hz). Anal. Calcd for $C_{12}H_{14}FNO_2$: C, 64.56; H, 6.32; F, 8.51; N, 6.28. Found: C, 64.52; H, 6.29; F, 8.62; N, 6.35.

N-Hydroxysuccinimide esters (24 and 25) of 15 and 19.—A mixture of 15 (or 19) (1.0 mmol), N-hydroxysuccinimide (1.02 mmol), and N,N'-dicyclohexylcarbodiimide (1.0 mmol) in dry EtOAc (5.5 mL) was stirred for 1 h at room temperature. The resultant precipitate was filtered off, washed with EtOAc, and the combined filtrate and washings were concentrated to give syrupy 24 (or 25), which showed a single spot at R_f 0.45 in TLC (4:1 toluene–EtOAc), and was used without further purification.

1-N-[(2S,4S)-5-Amino-4-fluoro-2-hydroxypentanoyl]dibekacin (26).—To a solution of 23 [13] $(716 \,\mathrm{mg}, \, 0.75 \,\mathrm{mmol}) \,\mathrm{in} \, 2:1 \,\mathrm{THF-H_2O} \,(40 \,\mathrm{mL}) \,\mathrm{was}$ added **24** (580 mg, \sim 1.6 mmol) in THF (10 mL) and, after the pH had been adjusted to ~8 by addition of aq NaHCO₃ (satd), the solution was kept for 1h at room temperature. Concentration gave a residue, which was washed with water and EtOAc. The solid obtained (632 mg) was dissolved in M NH₃ in 2:1 THF-H₂O (36 mL) and the solution was kept for 40 h at room temperature [de(trifluoroacetyl)ation]. Concentration gave a residue, which was dissolved in 30:15: 1 1,4-dioxane-H₂O-AcOH (28 mL) and hydrogenated in the presence of Pd-black for 5 h. After filtration, the filtrate was concentrated, and the solid was chromatographed on a CM Sephadex C-25 column (aq $0\rightarrow0.15\,\mathrm{M}$ NH₃) to give **26** as a solid, which was dried thoroughly in vacuo $(0.2\sim1 \text{ mmHg})$ in a desiccator for 3 days in the presence of P₂O₅ (225 mg, 43%); $[\alpha]_{D}^{23} + 71^{\circ} (c 1, H_{2}O); {}^{1}H NMR (DCl-D_{2}O, pD)$ 3): δ (signals relating to the side chain are shown mainly) 5.83 (d, 1 H, H-1'), 5.22 (d, 1 H, H-1"), 5.11 (double multiplets, 1 H, H-4"), 4.41 (dd, 1 H, H-2"'), 2.28 (dddd, 1 H, H-3"b), 1.93 (dddd, 1 H, H-3"'a); $J_{1',2'} \approx J_{1'',2''}$ 3.8, $J_{2''',3'''a}$ 11, $J_{2''',3'''b}$ 3, $J_{3'''a,3'''b}$ 15, $J_{3'''a,4'''}$ 2.5, $J_{3'''b,4'''}$ 10, $J_{3'''a,F}$ 37, $J_{3'''b,F}$ 13, $J_{4''',F} \sim 51$ Hz. ¹⁹F NMR (DCl-D₂O, Freon 11 as the external reference, pD 3): δ –192.5 (m). Anal. Calcd for $C_{23}H_{45}FN_6O_{10}\cdot 1.5$ H₂CO₃·H₂O: C, 42.30; H, 7.24; F, 2.73; N, 12.08. Found: C, 42.11; H, 7.30; F, 2.78; N, 11.98.

1-N-[(2S,4R)-5-Amino-4-fluoro-2-hydroxy-pentanoyl]dibekacin (27).—Compound 23 (530 mg, 0.56 mmol) was treated with 25 (410 mg, 1.1 mmol) in the manner described for 26 to give 27 as a solid (178 mg, 49%); $[\alpha]_D^{22} + 81^\circ$ (c 1, H₂O); ¹H NMR (DCl-D₂O, pD 3): δ 5.82 (d, 1 H, H-1'), 5.22 (d, 1 H, H-1"), 5.15 (double multiplets, 1 H, H-4"'), 4.42 (dd, 1 H, H-2"'), 2.25 (ddt, 1 H, H-3"'b), 2.18 (ddt, 1 H, H-3"'a); $J_{1',2'} \approx J_{1'',2''}$ 3.8, $J_{2''',3'''a}$ 7.5, $J_{2''',3'''b}$ 4.5, $J_{3'''a,3'''b}$ 15, $J_{3'''a,4'''}$ 7.5, $J_{3'''b,4'''}$ 4.5, $J_{3'''a,F}$ 18.5, $J_{3'''b,F}$ 30, $J_{4''',F} \sim 50$ Hz. ¹⁹F NMR (DCl-D₂O, Freon-11 as the external reference, pD 3): δ –190.3 (m). Anal. Calcd for $C_{23}H_{45}FN_6O_{10}\cdot H_2CO_3\cdot 0.5$ H₂O: C, 43.96; H, 7.38; F, 2.90; N, 12.82. Found: C, 43.94; H, 7.51; F, 3.04; N, 13.07.

1-N-[(S)-3-Amino-2-hydroxypropanoyl]dibekacin (28).—A mixture of (S)-N-(benzyloxycarbonyl)isoserine [15,16] (1.20 g, 5.02 mmol), N-hydroxysuccinimide (580 mg, 5.04 mmol), and N,N'-dicyclohexylcarbodiimide (1.04 g, 5.04 mmol) in THF (25 mL) was stirred for 2 h at room temperature. The precipitate was filtered off and washed thoroughly with THF. The combined filtrate and washings were concentrated to give the active ester of the protected isoserine as a syrup. Subsequently the syrup was treated with 23 (2.74 g, 2.88 mmol) as described for 26 to give a solid, which was subjected to chromatography on a column of CM-Sephadex C-25 (aq $0\rightarrow0.5$ M NH₃) to give **28** as a solid (920 mg, 53%), $[\alpha]_D^{25} + 89^\circ$ (c 1, H₂O) (lit [14], no data reported); ¹H NMR (DCl–D₂O, pD 3): δ 5.84 (d, 1 H, H-1'), 5.21 (d, 1 H, H-1"), 4.52 (dd, 1 H, H-2"), 3.44 (dd, 1 H, H-3"b), 3.28 (dd, 1 H, H-3"a), 2.32 (dt, 1 H, H-2eq), 1.94 (q, 1 H, H-2ax); $J_{1,2ax} \approx J_{2ax,2eq} \approx J_{2ax,3}$ 13, $J_{1,2eq} \approx J_{2eq,3}$ 4.5, $J_{1',2'} \approx J_{1'',2''}$ 3.8, $J_{2''',3'''a}$ 8.5, $J_{2''',3'''b}$ 4, $J_{3'''a,3'''b}$ 13 Hz. Anal. Calcd for $C_{21}H_{42}N_6O_{10}\cdot H_2CO_3$: C, 43.70; H, 8.00; N, 13.90. Found: C, 43.40; H, 7.68; N, 14.10.

1-N-[(S)-5-Amino-2-hydroxypentanoyl]dibekacin (29).—(S)-5-(Benzyloxycarbonyl)amino-2-hydroxypentanoic acid [15,17,18] (1.50 g, 5.61 mmol) was treated with N-hydroxysuccinimide (660 mg, 5.73 mmol) in the manner described for 24 to give the corresponding active ester as a syrup. Subsequently the syrup was treated with 23 (3.17 g,

3.34 mmol) as described for **28** to give **29** as a solid $(1.45 \text{ g}, 67\%); [\alpha]_D^{24} + 82^\circ (c \ 0.5, \text{ H}_2\text{O}); ^1\text{H NMR}$ (DCl–D₂O, pD 3): δ 5.83 (d, 1 H, H-1'), 5.22 (d, 1 H, H-1"), 4.20 (dd, 1 H, H-2""), 3.09 (slightly br t, 2 H, J 7.5 Hz, H-5"a,5"b), 2.29 (dt, 1 H, H-2eq), \sim 1.93 (H-3"b), 1.91 (q, 1 H, H-2ax), 1.77–1.95 (H-4'''a,4'''b), 1.62-1.74 (m, 2 H, H-4'ax, 3'''a); $J_{1',2'} \approx J_{1'',2''}$ 3.8, $J_{2''',3'''a}$ 9, $J_{2''',3'''b}$ 3.5 Hz. Anal. Calcd for C₂₃H₄₆N₆O₁₀·H₂CO₃·H₂O: C, 44.57; H, 7.79; N, 13.00. Found: C, 44.47; H, 7.92; N, 12.55. 1-N-[(S)-6-Amino-2-hydroxyhexanoyl]dibekacin (30).—(S)-6-(Benzyloxycarbonyl)amino-2-hydroxyhexanoic acid [15,18] (1.50 g, 5.33 mmol) was treated with *N*-hydroxysuccinimide $(625 \, \text{mg},$ 5.43 mmol) in the manner described for **24** to give the active ester as a syrup. Subsequently the syrup was treated with 23 (3.20 g, 3.37 mmol) as described for **28** to give **30** as a solid (1.40 g, 63%), $[\alpha]_{D}^{25}$ $+80^{\circ}$ (c 1, H₂O); ¹H NMR (DCl–D₂O, pD 3): δ 5.85 (d, 1 H, H-1'), 5.24 (d, 1 H, H-1"), 4.20 (dd, 1 H, H-2"'), 3.07 (slightly br t, 2 H, J 7.5 Hz, H-6"a,6"b), 2.31 (dt, 1 H, H-2eq), 1.93 (q, 1 H, H-2ax), 1.88 (dddd, 1 H, H-3"b), 1.76 (m, 2 H, H-5'''a,5'''b), ~1.68 (H-3'''a), 1.55 (m, 2 H, H- $4'''a,4'''b); J_{1'.2'} \approx J_{1'',2''} 3.8, J_{2''',3'''a} 9, J_{2''',3'''b} 4,$ $J_{3'''a,3'''b}$ 14, $J_{3'''b,4'''a}$ 6.5 (or 10), $J_{3'''b,4'''b}$ 10 (or Anal. Calcd for $C_{24}H_{48}N_6O_{10}\cdot H_{2-}$ CO₃·H₂O: C, 45.45; H, 7.93; N, 12.72. Found: C, 45.71; H, 7.73; N, 12.52.

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